



# *FUM* Gene Expression Profile and Fumonisin Production by *Fusarium verticillioides* Inoculated in *Bt* and Non-*Bt* Maize

Liliana O. Rocha<sup>1\*</sup>, Vinicius M. Barroso<sup>1</sup>, Ludmila J. Andrade<sup>1</sup>, Gustavo H. A. Pereira<sup>2</sup>, Fabiane L. Ferreira-Castro<sup>1</sup>, Aildson P. Duarte<sup>3</sup>, Marcos D. Michelotto and Benedito Correa<sup>1</sup>

<sup>1</sup> Laboratório de Micotoxinas, Departamento de Microbiologia, Instituto de Ciências Biomédicas, Universidade de São Paulo, São Paulo, Brazil, <sup>2</sup> Departamento de Estatística, Centro de Ciências Exatas e de Tecnologia, Universidade Federal de São Carlos, São Carlos, Brazil, <sup>3</sup> Centro de Grãos e Fibras, Instituto Agrônomo de Campinas, Agência Paulista de Tecnologia dos Agronegócios (APTA), Campinas, Brazil

## OPEN ACCESS

### Edited by:

Andrea Gomez-Zavaglia,  
Center for Research  
and Development in Food  
Cryotechnology (CIDCA, CONICET),  
Argentina

### Reviewed by:

Valeria Scala,  
Sapienza University of Rome, Italy  
Agnieszka Waskiewicz,  
Poznan University of Life Sciences,  
Poland

### \*Correspondence:

Liliana O. Rocha  
lilianarochoa@usp.br

### Specialty section:

This article was submitted to  
Food Microbiology,  
a section of the journal  
Frontiers in Microbiology

**Received:** 08 November 2015

**Accepted:** 14 December 2015

**Published:** 06 January 2016

### Citation:

Rocha LO, Barroso VM, Andrade LJ,  
Pereira GHA, Ferreira-Castro FL,  
Duarte AP, Michelotto MD  
and Correa B (2016) *FUM* Gene  
Expression Profile and Fumonisin  
Production by *Fusarium verticillioides*  
Inoculated in *Bt* and Non-*Bt* Maize.  
*Front. Microbiol.* 6:1503.  
doi: 10.3389/fmicb.2015.01503

This study aimed to determine the levels of fumonisins produced by *Fusarium verticillioides* and *FUM* gene expression on *Bt* (*Bacillus thuringiensis*) and non-*Bt* maize, post harvest, during different periods of incubation. Transgenic hybrids 30F35 YG, 2B710 Hx and their isogenic (30F35 and 2B710) were collected from the field and a subset of 30 samples selected for the experiments. Maize samples were sterilized by gamma radiation at a dose of 20 kGy. Samples were then inoculated with *F. verticillioides* and analyzed under controlled conditions of temperature and relative humidity for fumonisin B<sub>1</sub> and B<sub>2</sub> (FB<sub>1</sub> and FB<sub>2</sub>) production and *FUM1*, *FUM3*, *FUM6*, *FUM7*, *FUM8*, *FUM13*, *FUM14*, *FUM15*, and *FUM19* expression. 2B710 Hx and 30F35 YG kernel samples were virtually intact when compared to the non-*Bt* hybrids that came from the field. Statistical analysis showed that FB<sub>1</sub> production was significantly lower in 30F35 YG and 2B710 Hx than in the 30F35 and 2B710 hybrids ( $P < 0.05$ ). However, there was no statistical difference for FB<sub>2</sub> production ( $P > 0.05$ ). The kernel injuries observed in the non-*Bt* samples have possibly facilitated *F. verticillioides* penetration and promoted FB<sub>1</sub> production under controlled conditions. *FUM* genes were expressed by *F. verticillioides* in all of the samples. However, there was indication of lower expression of a few *FUM* genes in the *Bt* hybrids; and a weak association between FB<sub>1</sub> production and the relative expression of some of the *FUM* genes were observed in the 30F35 YG hybrid.

**Keywords:** transgenic corn, mycotoxins, *Fusarium*, gene expression, *Bacillus thuringiensis*

## INTRODUCTION

Maize (*Zea mays* L.) is one of the world's major agricultural crops serving as a staple food for millions, with an annual average production and consumption of 989.2 million metric tons (Mt) and 953.9 Mt (2013-2014), respectively. Brazil is currently the world's third largest producer, after United States and China, with an average production of 79.3 Mt over the last year (United States Department of Agriculture [USDA], 2014).

Susceptible to fungal and mycotoxin contamination, numerous toxic fungal secondary metabolites can be found in maize, though, fumonisins occur with greatest frequency. Members of the *Fusarium fujikuroi* species complex, mainly *F. verticillioides* (Sacc.) Nirenberg and *F. proliferatum* (Matsush.) Nirenberg, are capable of producing these mycotoxins consistently (Desjardins, 2006). *F. verticillioides* is one of the most important species associated with maize throughout the world and produces high levels of fumonisins. Contamination with these mycotoxins has a significant impact, compromising the quality of maize products (de la Campa et al., 2005).

Based on chemical structure, fumonisins have been classified into A, B, C, and P groups; the most common are the B analogs and fumonisin B<sub>1</sub> (FB<sub>1</sub>) is the most prevalent and toxic of the group (Stepien et al., 2011). Fumonisin B<sub>1</sub> is known to cause toxicity in animals and humans due to the inhibition of sphingolipid metabolism and cell cycle regulation, resulting in diverse and complex effects. Examples include leukoencephalomalacia in horses, pulmonary oedema, and hydrothorax in swine; fumonisin B<sub>1</sub> is additionally carcinogenic to rodents and exhibits nephrotoxic as well as hepatotoxic activity in rats and rabbits (Desjardins, 2006). Epidemiological studies have suggested that fumonisins may also be associated with oesophageal cancer and neural tube birth defects in humans (Marasas et al., 2004).

In *F. verticillioides*, fumonisin production is regulated by the fumonisin biosynthetic gene cluster (*FUM*), which consists of 16 genes encoding biosynthetic enzymes, regulatory, and transport proteins (Proctor et al., 2013). *FUM1* catalyzes the synthesis of a linear polyketide that forms the backbone structure of fumonisins. *FUM8* encodes an  $\alpha$ -oxoamine synthase responsible for the condensation of the linear polyketide with alanine (Proctor et al., 2003, 2013; Lazzaro et al., 2012; Medina et al., 2013). The cluster also encodes a C-3 carbonyl reductase (*FUM13*), and cytochrome P450 oxygenases (*FUM2*, *FUM3*, *FUM6*, *FUM15*; Proctor et al., 2003; Bojja et al., 2004). The genes *FUM7*, *FUM10*, *FUM11*, *FUM14*, and *FUM16* are required for tricarballic acid esterification. *FUM17* and *FUM18* are longevity assurance factors; and *FUM19* encodes a protein highly similar to ABC multidrug transporters, which can possibly reduce cellular concentration of toxins, therefore conferring self-protection (Proctor et al., 2003; Bojja et al., 2004; Desjardins, 2006). *FUM21* encodes a Zn(II)<sub>2</sub>Cys<sub>6</sub> binuclear DNA-binding transcription factor that positively regulates *FUM* gene expression (Brown et al., 2007). Studies have demonstrated that these genes are correlated with fumonisin production (Lopez-Errasquin et al., 2007; Medina and Magan, 2010; Rocha et al., 2011) and the expression of some mycotoxin genes respond positively to different environmental conditions (Medina and Magan, 2010).

Globally, fumonisin contamination in food is a concern. The FDA (the United States Food and Drug Administration) has established a limit of 3–4 ppm of fumonisin contamination (FB<sub>1</sub> + FB<sub>2</sub> + FB<sub>3</sub>) in human foods and 5–100 ppm in animal feeds. The Commission Regulation (EC) of the European Union has established a maximum level of 4 ppm for FB<sub>1</sub> + FB<sub>2</sub> in unprocessed maize and 1 ppm in maize

and maize-based products intended for human consumption (European Commission Regulation [EC], 2007). However, the legislation is distinct around the world, depending on the concern that the authorities have about the potential toxic effects of fumonisins on animals and their implications for industry (Food and Drug Administration [FDA], 2001; Schatzmayr and Streit, 2013).

Low levels of fumonisins can occur even in intact maize kernels, since *F. verticillioides* is found in both asymptomatic and diseased plants. Environmental conditions, water availability and the genetic background of the plant and the pathogen are significant factors in disease development and mycotoxin production (Oren et al., 2003; Bowers et al., 2013). It has been shown that physically injured kernels increase the propensity for fungal contamination and mycotoxin production (Munkvold et al., 1999; Parsons and Munkvold, 2012; Bowers et al., 2013). Kernel injuries caused by ear feeding insects are particularly important. These pests act as vectors for fungal spores, increasing the severity of fungal disease and mycotoxin production (Dowd, 2000; Wu, 2006; Ferreira-Castro et al., 2012).

The damage caused by European corn borer (ECB, *Ostrinia nubilalis* Hübner), Southwestern corn borer (SWCB, *Diatraea grandiosella* Dyar), corn earworm (CEW, *Helicoverpa zea* Boddie) and fall armyworm (FAW, *Spodoptera frugiperda*, J.E. Smith) has been shown to favor mycotoxin contamination in the field and to contribute to mycotoxin accumulation during storage (Sinha, 1994; Dowd, 2000; Wu, 2006). *Bt* (*Bacillus thuringiensis*) hybrids have been effective in reducing injuries by these insects, indirectly controlling plant susceptibility to fungal infection and mycotoxin contamination (Munkvold et al., 1997; Munkvold and Hellmich, 1999; Ostry et al., 2010; Bowers et al., 2013; Foresti et al., 2013).

It is widely known that *Bt* maize presents a lower risk of fumonisin contamination compared with non-*Bt* hybrids when exposed to lepidopteran insects (Duvick, 2001; Pazzi et al., 2006; Abbas et al., 2011; Bowers et al., 2013, 2014). A recent study has also demonstrated that *Bt* maize exhibited reduction in deoxynivalenol and zearalenone contamination in harvested maize kernels (Ostry et al., 2010). Although many studies have shown the relationship between kernel injuries by insects and fumonisin contamination in maize during harvest, none has focused on the fumonisin accumulation in *Bt* and non-*Bt* maize after this period.

The aim of this study was to verify the levels of fumonisins produced by *F. verticillioides* on *Bt* and non-*Bt* maize, post harvest, in different periods of incubation under controlled conditions. The second objective of this study was to compare *FUM* gene expression between *Bt* and non-*Bt* hybrids and to study the association between fumonisin production and *FUM* gene expression by *F. verticillioides* for each of the hybrids.

## MATERIALS AND METHODS

### Maize Grain Samples

Maize cultivar samples of *Bt* 2B710 Hx and 30F35 YG and their isogenic non-*Bt* 2B710 and 30F35 were provided by the

Agronomic Institute of Campinas (Instituto Agronômico de Campinas, APTA-São Paulo, Brazil). These samples were sown in November/2010 and harvested in March/2011 in Cruzália, State of São Paulo, Brazil.

Sampling was conducted according to methodology proposed by Delp et al. (1986) with modifications. *Bt* and non-*Bt* crop fields were sampled by dividing them into four sectors of uniform size containing eight rows each. Ten samples were randomly collected from each sector, for a total of 40 samples per hybrid. The hybrids were harvested manually in the experimental field by the technical support staff of the Agronomic Institute of Campinas. Thirty subsamples for each hybrid were then selected for this experiment, totalling 120 tests.

Ten samples of 30F35 YG (expressing Cry 1Ab protein, equivalent to MON810, Monsanto), 10 samples of 2B710 Hx (expressing Cry 1F protein, equivalent to TC1 507, Dow Agrosciences) corn grains and their respective isogenic hybrids were used for each incubation period (10, 20, and 30 days).

The injuries caused by *S. frugiperda* in 30F35 YG, 2B710 Hx and their isogenic hybrids were previously analyzed from sowing to harvest by The Agronomic Institute of Campinas in the same experimental field used in the current study (Michelotto et al., 2011).

All samples were sterilized by gamma radiation at a dose of 20 kGy to eliminate the natural mycoflora at the Nuclear and Energetic Research Institute (Instituto de Pesquisas Energéticas e Nucleares-IPEN-CNEN/SP). A GammaCell irradiator equipped with a cobalt 60 source was used and a dose rate ranging from 4.74 to 4.84 kGy/h was applied. The mycoflora was verified in order to control the irradiation procedure (Berjak, 1984). Samples were previously analyzed for the presence of fumonisins according to the methodology described by Visconti et al. (2001), because gamma radiation does not eliminate fumonisin from grains (Ferreira et al., 2007). The values (for each sample) were considered to be a basal FB<sub>1</sub> or FB<sub>2</sub> contamination, and were used to calculate the final fumonisin concentration produced by *F. verticillioides* after each period of incubation (total concentration minus basal concentration).

## Fusarium verticillioides Strain

*Fusarium verticillioides* isolate 13BA was obtained from the culture collection of the Institute of Biomedical Sciences, ICBII, University of São Paulo, Brazil. The isolate was maintained on SNA (Spezieller Nährstoffarmer Agar) by monospore isolation and it was used in all experiments (Rocha et al., 2011).

Before beginning the experiments, the ability of fumonisin production by *F. verticillioides* strain 13BA was re-evaluated according to methodology proposed by Ross et al. (1990) and Sydenham et al. (1990), with concentrations of 120 and 36 µg/g for FB<sub>1</sub> and FB<sub>2</sub>, respectively.

## Inoculation of Corn Samples with Fusarium verticillioides Spore Suspension

The *F. verticillioides* fungal spore suspension was prepared according to Ferreira et al. (2007). The final suspension was

adjusted to  $5 \times 10^5$  spores/mL and inoculated into 5 g of maize bran (previously treated with 20 kGy gamma radiation). This mixture was added to Petri dishes containing 25 g of grains used for fumonisin and *FUM* gene cluster expression analyses. The samples were stored in a plastic container at a relative humidity and temperature controlled by a previously calibrated thermo hygrometer. The container was incubated in a BOD incubator at 25°C and a relative humidity of 97.5 to 99.0% obtained with 30% K<sub>2</sub>SO<sub>4</sub> solution as described by Winston and Bates (1960) for 10, 20, and 30 days. After each incubation period, 10 samples of each hybrid were analyzed, with 20 g for fumonisin analyses and 5 g for gene expression experiments.

## Determination of Fumonisins

Samples were prepared and extracted according to the methodology proposed by Visconti et al. (2001). Afterwards, fumonisins were mixed with the derivatizing agent ortho-phthalaldehyde (OPA; Visconti et al., 2001) and injected into a Shimadzu liquid chromatograph (LC-10AD). Separation was performed on a C-18 reverse phase column (5 ODS-20, 150 mm × 4.6 mm; Phenomenex).

Fumonisins were quantified based on a calibration curve using standard solutions of FB<sub>1</sub> and FB<sub>2</sub> (Sigma-Aldrich). The coefficient of correlation was 0.993 for FB<sub>1</sub> and 0.995 for FB<sub>2</sub>. The limit of quantification (LOQ) for FB<sub>1</sub> was 0.015 µg/g, with a mean recovery of 92.38% and standard deviation of 13.68% (five replicates). For FB<sub>2</sub>, LOQ was 0.015 µg/g, with a mean recovery of 85.39% and standard deviation of 6.87% (five replicates). LOQ was determined via recovery tests. The lowest validated spike (0.015 µg/g) level with a percentage of recovery between 70 and 120% and a standard variation lower than 20% was defined as the method LOQ in this study (European Commission Regulation [EC], 2007).

The final fumonisin concentration was calculated through the subtraction of the total FB<sub>1</sub> or FB<sub>2</sub> for each analyzed sample (basal FB<sub>1</sub> or FB<sub>2</sub> concentration plus the concentration of FB<sub>1</sub> or FB<sub>2</sub> produced by *F. verticillioides* during the respective incubation periods) minus the basal FB<sub>1</sub> or FB<sub>2</sub> contamination of each sample that came from the field.

## FUM Gene Cluster Expression

### Isolation of mRNA and Reverse Transcription

RNA was extracted from 5 g of corn samples corresponding to each inoculation period (10, 20, and 30 days) using the Plant RNA Isolation Aid Kit (Ambion®, Life Technologies, Carlsbad, CA, USA) followed by purification utilizing RNAqueous Kit (Ambion®, Life Technologies, Carlsbad, CA, USA), according to the manufacturer's instructions. cDNA was synthesized with the High Capacity cDNA Reverse Transcription kit according to the manufacturer's instructions (Applied Biosystems, Life Technologies, Carlsbad, CA, USA). cDNA was synthesized in a Gene Amp® PCR System 9700 thermal cycler (Applied Biosystems, Life Technologies) for 1 h at 37°C. The samples were stored at -20°C.

## Quantitative PCR (qPCR)

In this study, the following *FUM* genes were initially tested: *FUM1*, *FUM2*, *FUM3*, *FUM6*, *FUM7*, *FUM8*, *FUM10*, *FUM13*, *FUM14*, *FUM15*, *FUM19*, and *FUM21*. Primers were designed in the program Primer 3<sup>1</sup> from the reference sequences AF155773 (Proctor et al., 1999) and U27303/EU430619 (Yan and Dickman, 1996; Visentin et al., 2009) deposited at the National Centre for Biotechnology Information-NCBI<sup>2</sup>. *FUM1*, *FUM19*, and *TUB* primers were obtained from the study of Lopez-Errasquín et al. (2007; **Table 1**). The specificity of the primer sequences was verified through BLAST tool in NCBI.

The Platinum SYBR Green qPCR SuperMix-UDG reagent (Invitrogen, Life Technologies) was used as the reaction mixture, according to the manufacturer's instructions. Each sample was analyzed in duplicate. An appropriate negative control containing no template was used in all reactions to exclude possible contaminations. The quantification of mRNA was normalized using the average of *TUB* ( $\beta$ -tubulin) and *CALM* (calmodulin) genes (Bustin et al., 2009), with cDNA amplifications ran on the same plate. The following reference genes were tested prior to the experiments: *CALM*, *TUB*, *EF-1 $\alpha$*  (elongation factor 1  $\alpha$ ) and *18s rRNA* (18s ribosomal RNA). The Ct values of *TUB* and *CALM* were similar; therefore, the mean values of these genes were used for normalizing the data.

Relative quantification based on  $\Delta\Delta C_t$  values was the method of analysis used in this study, since the efficiencies of

compatibility tests between the average of the reference genes (*TUB* and *CALM*) and the target gene were similar, with slopes between  $-0.1$  and  $0.1$  (Pfaffl, 2001; Gizinger, 2002; Schmittgen and Livak, 2008; Raymaekers et al., 2009).

For validation of the method, serial dilutions of the target genes and of the reference genes were prepared in triplicate. The primers' efficiency was calculated based on the curve generated by the software. *FUM2*, *FUM10*, and *FUM21* were excluded from the analysis since qPCR parameters were not acceptable according to the method of analysis proposed by Pfaffl (2001) and Schmittgen and Livak (2008).

## Statistical Analysis

The results were analyzed using Gamlss R 2.9 package and Statistical Analysis Software (SAS) version 9.1. The Gamlss model (5% level of significance) with Weibull distribution was used to evaluate differences of FB<sub>1</sub> and FB<sub>2</sub> production between *Bt* and non-*Bt* maize during the period of incubation (10, 20, and 30 days; Rigby and Stasinopoulos, 2005). The Weibull distribution was chosen using quantile residual plots (Dunn and Smyth, 1996). The analysis was conducted based on the overall data (from the first to 30th day). The Gamlss model was used to verify the interaction between hybrids (*Bt* and non-*Bt*) and period of incubation (10, 20, and 30 days). As these interactions were not significant ( $P > 0.05$ ), further analyses based on each period of incubation (10, 20, and 30 days) were not conducted.

The differences of *FUM* gene expression between *Bt* and non-*Bt* maize were evaluated using the Mann-Whitney test, since the data was not normally distributed (5% level of significance)

<sup>1</sup><http://frodo.wi.mit.edu/primer3/>

<sup>2</sup><http://www.ncbi.nlm.nih.gov/>

**TABLE 1 | qPCR primers used to evaluate *FUM* gene expression by *Fusarium verticillioides* in maize grains.**

Locus	Primer sequence (5'-3')	Fragment size (bp)	Reference/NCBI accession number
<i>FUM1F</i> <i>FUM1R</i>	GAGCCGAGTCAGCAAGGATT AGGGTTCGTGAGCCAAGGA	90	Lopez-Errasquín et al., 2007
<i>FUM3F</i> <i>FUM3R</i>	CTTGCGGTGCCCATACTA GGACCAAGAGCGTGATG	60	This study/AF155773
<i>FUM6F</i> <i>FUM6R</i>	GATAGACTCGGGGCTGAGA AGCTCGCCGACAGAATC	100	This study/AF155773
<i>FUM7F</i> <i>FUM7R</i>	CATCGTATCTACATTGTCGCATC TGTA CTCTCCAACAATATGAATGAGTC	100	This study/AF155773
<i>FUM8F</i> <i>FUM8R</i>	CAACAGAAATACGCAATGACG TGCTCGACCACTACATCAGG	99	This study/AF155773
<i>FUM13F</i> <i>FUM13R</i>	GCCTTTGGTCTTGTCTCTCA CGTCAATTATTGCCTCTTTCAA	100	This study/AF155773
<i>FUM14F</i> <i>FUM14R</i>	TAGGTCCAGGTCGAGATGCT GGAAGCCAAGAACCCAATCT	99	This study/AF155773
<i>FUM15F</i> <i>FUM15R</i>	TGCCATCCAGAATGACGATA GAGTCTCAGGAGAGCGAGGA	94	This study/AF155773
<i>FUM19F</i> <i>FUM19R</i>	ATCAGCATCGGTAACGCTTATGA CGCTTGAAGAGCTCCTGGAT	88	Lopez-Errasquín et al., 2007
<i>TUBF</i> <i>TUBR</i>	CCGGTATGGGTACTCTGCTC CTCAACGACGGTGTGACAGAA	95	Lopez-Errasquín et al., 2007
<i>CALMF</i> <i>CALMR</i>	ACGGTTTCATTCTGCTGCT TCAGCCTCTCGGATCATCTC	97	This study/AF155773



for each period of incubation (Gibbons and Chakraborti, 2003). Pearson's correlation tests were performed to investigate possible associations between the relative expression levels of *FUM1*, *FUM3*, *FUM6*, *FUM7*, *FUM8*, *FUM13*, *FUM14*, *FUM15*, and *FUM19* and the production of FB<sub>1</sub> and FB<sub>2</sub> by *F. verticillioides* on *Bt* and non-*Bt* samples.

## RESULTS

### Production of FB<sub>1</sub> and FB<sub>2</sub> in *Bt* and Non-*Bt* Maize Samples Artificially Contaminated with *F. verticillioides* During the Periods of 10, 20, and 30 days

FB<sub>1</sub> and FB<sub>2</sub> were quantified during 10, 20, and 30 days of incubation. The *Bt* hybrid 30F35 YG presented FB<sub>1</sub> levels between 0.11 to 4.99 µg/g (mean: 2.38 µg/g) and FB<sub>2</sub> from 0.02 to 2.66 µg/g (mean: 1.53 µg/g). 30F35 hybrid presented FB<sub>1</sub> levels from 1.49 to 5.34 µg/g (mean: 3.17 µg/g) and FB<sub>2</sub> from 0.015 to 2.83 µg/g (mean: 1.7 µg/g) (Table 2). Statistical analysis showed that FB<sub>1</sub> contamination was significantly lower in 30F35YG when compared to its isogenic hybrid 30F35 ( $P = 0.0095$ ). However, there was no statistical difference between *Bt* and non-*Bt* maize samples for FB<sub>2</sub> production ( $P = 0.827$ ). Interaction between hybrids (*Bt* and non-*Bt*) and period of incubation (10, 20, and 30 days) was not significant ( $P > 0.05$ ), therefore there was no evidence that the mean difference of FB<sub>1</sub> ( $P = 0.26$ ) and FB<sub>2</sub> ( $P = 0.82$ ) production between *Bt* and non-*Bt* maize changed over the period of incubation.

2B710 Hx presented levels of FB<sub>1</sub> and FB<sub>2</sub> between 0.19–9.71 µg/g (mean: 6.42 µg/g) and 0.02–6.75 µg/g (mean: 3.98 µg/g), respectively. The isogenic 2B710 hybrid presented FB<sub>1</sub> levels from 3.33–11.55 µg/g (mean: 8.49 µg/g) and FB<sub>2</sub> from 0.8 to 8.31 µg/g (mean: 4.67 µg/g) (Table 2). FB<sub>1</sub> contamination was significantly lower in 2B710Hx when compared to its isogenic hybrid 2B710 ( $P = 0.0001$ ). There was no statistical difference between *Bt* and non-*Bt* maize samples for FB<sub>2</sub> production ( $P = 0.382$ ). There was no evidence that the mean difference of FB<sub>1</sub> ( $P = 0.95$ ) and FB<sub>2</sub> ( $P = 0.61$ ) production between *Bt* and non-*Bt* maize changed over the period of incubation.

### Optimization of qPCR Reactions and *FUM* Gene Expression in *Bt* and Non-*Bt* Maize Samples Artificially Contaminated with *F. verticillioides* During the Periods of 10, 20, and 30 days

The comparative  $\Delta\Delta C_t$  method was chosen for *FUM* gene expression analysis, since qPCR efficiencies were similar.  $\Delta\Delta C_t$  relative quantification is used to compare the gene expression between one sample and another (control). In this study, the control was considered the samples with the lowest  $C_t$  value for all of the genes for each hybrid (Supplementary Tables S1 and S2).

The efficiencies ( $E$ ) obtained in the qPCR analysis were between 78 and 110% for all of the studied genes, with slopes

between  $-4$  (*FUM21*) and  $-3.1$  (*FUM8*, *FUM14*, *FUM15*) and  $r^2$  from 0.97 (*FUM2*, *FUM6*, *FUM10*) to 0.99 (all the other genes; Supplementary Figure S1). According to the European Network of GMO Laboratories, acceptable slopes for standard curve range from  $-3.1$  to  $-3.6$ . In this study, *FUM2*, *FUM10*, and *FUM21* presented values outside this range (European Network of Gmo Laboratories [ENGL], 2008). The melt-curve analysis resulted in one single peak for all of the genes used in the analysis (Supplementary Figure S1).

The average between the  $C_t$  values of the reference genes  $\beta$ -tubulin (*TUB*) and *CALM* were selected to normalize the  $C_t$  values of the samples. The average of *CALM* and *TUB* showed better repeatability and less variability in  $C_t$  values when compared to other tested genes (*EF-1 $\alpha$*  and *18s rRNA*). The  $\Delta C_t$  values were plotted against the log dilutions, and the slopes ranged from  $-0.24$  (*FUM21* in relation to *TUB* and *CALM*) to  $0.02$  (*FUM1* in relation to *TUB* and *CALM*). *FUM2*, *FUM10*, and *FUM21* reactions did not present acceptable validation parameters once slope values for compatibility tests were not within  $-0.1$  and  $0.1$ ; therefore they were excluded from the analysis. Even though  $r^2$  for *FUM6* was below 0.99, this gene was not excluded from the analyses, since efficiency and slope values were satisfactory.

Amplification of the *FUM* genes was observed in all of the samples. Statistical analysis showed that there was an indication of higher *FUM1* expression during the three periods of incubation ( $P < 0.02$ ), higher *FUM13* expression during the 10th day of incubation and higher *FUM7*, *FUM8*, and *FUM14* expressions during the 30th day of incubation ( $P < 0.04$ ) and higher levels of *FUM19* expression in the 20 and 30th days of incubation ( $P < 0.04$ ) in the non-*Bt* 30F35 hybrid. There was an indication of higher *FUM19* expression during the 10th day of incubation in the non-*Bt* 2B710 hybrid ( $P < 0.03$ ).

There was a weak correlation between FB<sub>1</sub> production and the expression of *FUM1* ( $P = 0.009$ ,  $r = 0.47$ ), *FUM7* ( $P = 0.017$ ,  $r = 0.43$ ), *FUM8* ( $P = 0.024$ ,  $r = 0.41$ ), *FUM13* ( $P = 0.032$ ,  $r = 0.39$ ), *FUM14* ( $P = 0.001$ ,  $r = 0.57$ ), *FUM15* ( $P = 0.011$ ,  $r = 0.46$ ), and *FUM19* ( $P = 0.009$ ,  $r = 0.47$ ) in corn samples of 30F35 YG hybrid at a 5% significance level. For this hybrid, *FUM19* was also slightly correlated to FB<sub>2</sub> production ( $P = 0.049$ ,  $r = 0.36$ ). However, there is no evidence of correlation between FB<sub>1</sub> and FB<sub>2</sub> production and the studied *FUM* genes for the hybrids 30F35, 2B710, and 2B710 Hx (Figures 1 and 2).

## DISCUSSION

Samples representing the non-*Bt* hybrids 2B710 and 30F35 that came from the field presented damaged kernels as a result of insect injuries, which could possibly facilitate fungal penetration and consequently fumonisin production. 2B710 Hx and 30F35 YG samples were virtually intact (Michelotto et al., 2011). It has been shown that wounded plants and seeds facilitate fungal contamination, increasing the chance of diseases and mycotoxin production (Wallin, 1986; Gradziel and Wang, 1994; Mahoney and Rodrigues, 1996; Imathiu et al., 2008; Mohammadi and Hokmabadi, 2010).

**TABLE 2 | Fumonisin production by *F. verticillioides* in 30 maize samples of 30F35 YG, 2B710 Hx (*Bt*), 30F35 and 2B710 (non-*Bt* hybrids) during 10, 20, and 30 days of incubation under temperature of 25°C and humidity of 97.5 to 99.0%.**

Period of Incubation	Samples	30F35 YG ( <i>Bt</i> )		30F35 (non- <i>Bt</i> )		2B710 Hx ( <i>Bt</i> )		2B710 (non- <i>Bt</i> )	
		FB <sub>1</sub> (μg/g)	FB <sub>2</sub> (μg/g)	FB <sub>1</sub> (μg/g)	FB <sub>2</sub> (μg/g)	FB <sub>1</sub> (μg/g)	FB <sub>2</sub> (μg/g)	FB <sub>1</sub> (μg/g)	FB <sub>2</sub> (μg/g)
10 days	1	2,92 <sup>b</sup>	2,26	3,14	2,66	7,65	3,50	8,05	4,59
	2	2,91	2,60 <sup>b</sup>	3,78	2,83 <sup>b</sup>	6,72	2,61	10,90	5,68
	3	0,99	0,52	1,58	0,63	7,52	3,48	9,36	5,15
	4	1,99	0,60	2,07	0,41 <sup>a</sup>	0,19 <sup>a</sup>	0,02 <sup>a</sup>	7,92	3,68
	5	1,29	2,54	4,64 <sup>b</sup>	2,29	8,15	3,83	6,95	3,56
	6	0,32 <sup>a</sup>	0,61	2,06	0,78	6,01	6,03	8,98	2,59
	7	0,84	0,37 <sup>a</sup>	3,51	1,77	3,58	1,28	6,13	3,11
	8	2,02	0,57	4,19	2,21	7,70	5,68	4,46 <sup>a</sup>	2,89
	9	1,24	0,49	1,49 <sup>a</sup>	0,53	4,34	3,64	4,90	0,80 <sup>a</sup>
	10	2,64	2,29	2,71	2,30	8,24 <sup>b</sup>	6,75 <sup>b</sup>	11,03 <sup>b</sup>	8,31 <sup>b</sup>
Mean		<sup>c</sup> 1.72 ± 0.29*	1.29 ± 0.31	<sup>d</sup> 2.92 ± 0.35	1.64 ± 0.30	<sup>e</sup> 6.01 ± 0.82	3.68 ± 0.66	<sup>f</sup> 7.87 ± 0.72	4.04 ± 0.65
20 days	11	2,71	1,96	2,86	1,99	8,74	6,56	9,57	5,42
	12	4,99 <sup>b</sup>	1,64	5,11 <sup>b</sup>	1,64	7,26	5,89	10,34	5,87
	13	4,31	2,66 <sup>b</sup>	4,42	2,66 <sup>b</sup>	8,49	3,02	9,16	4,36
	14	3,47	2,01	3,76	2,03	7,24	3,68	10,97 <sup>b</sup>	6,58 <sup>b</sup>
	15	3,62	1,03	4,09	1,07	5,72	2,95	7,52	5,46
	16	3,48	1,40	3,65	1,41	6,11	2,24 <sup>a</sup>	5,05 <sup>a</sup>	2,46 <sup>a</sup>
	17	1,00	1,66	3,12	2,05	7,90	6,10	10,16	5,88
	18	3,03	2,27	3,16	2,27	4,97 <sup>a</sup>	3,05	6,83	3,45
	19	0,41 <sup>a</sup>	1,17	2,15	1,67	9,00 <sup>b</sup>	6,72 <sup>b</sup>	10,47	5,22
	20	1,00	0,02 <sup>a</sup>	1,84 <sup>a</sup>	0,015 <sup>a</sup>	5,64	3,98	7,37	4,45
Mean		<sup>c</sup> 2.8 ± 0.48	1.58 ± 0.23	<sup>d</sup> 3.42 ± 0.32	1.68 ± 0.23	<sup>e</sup> 7.11 ± 0.45	4.42 ± 0.54	<sup>f</sup> 8.74 ± 0.61	4.92 ± 0.39
30 days	21	2,87	1,55	2,96	1,55	4,43	3,64	9,24	7,45
	22	0,11 <sup>a</sup>	1,60	2,55	1,61	5,55	2,42	9,00	6,32
	23	2,30	0,91 <sup>a</sup>	2,45 <sup>a</sup>	0,92 <sup>a</sup>	5,56	1,55	9,43	7,90 <sup>b</sup>
	24	2,89	2,17 <sup>b</sup>	2,98	2,17	3,44	2,76	3,33 <sup>a</sup>	0,98 <sup>a</sup>
	25	2,83	1,88	2,90	1,88	8,43	4,95	11,55 <sup>b</sup>	7,47
	26	2,71	1,86	2,82	1,87	9,47	5,49	9,70	3,35
	27	3,72 <sup>b</sup>	2,06	4,01	2,08	9,71 <sup>b</sup>	6,62	7,86	3,62
	28	2,38	1,51	2,64	1,51	3,72	2,92	9,61	5,21
	29	2,96	1,83	3,15	1,83	2,59 <sup>a</sup>	1,50 <sup>a</sup>	8,22	2,56
	30	3,49	1,70	5,34 <sup>b</sup>	2,28 <sup>b</sup>	8,63	6,64 <sup>b</sup>	10,73	5,66
Mean		<sup>c</sup> 2.63 ± 0.31	1.71 ± 0.11	<sup>d</sup> 3.18 ± 0.28	1.77 ± 0.12	<sup>e</sup> 6.15 ± 0.85	3.85 ± 0.62	<sup>f</sup> 8.87 ± 0.70	5.05 ± 0.74
Mean/total		<sup>c</sup> 2.38 ± 0.22	1.53 ± 0.13	<sup>d</sup> 3.17 ± 0.18	1.7 ± 0.13	<sup>e</sup> 6.58 ± 0.41	3.98 ± 0.36	<sup>f</sup> 8.68 ± 0.31	4.67 ± 0.35

Fumonisin levels were calculated as described in section "Materials and Methods."

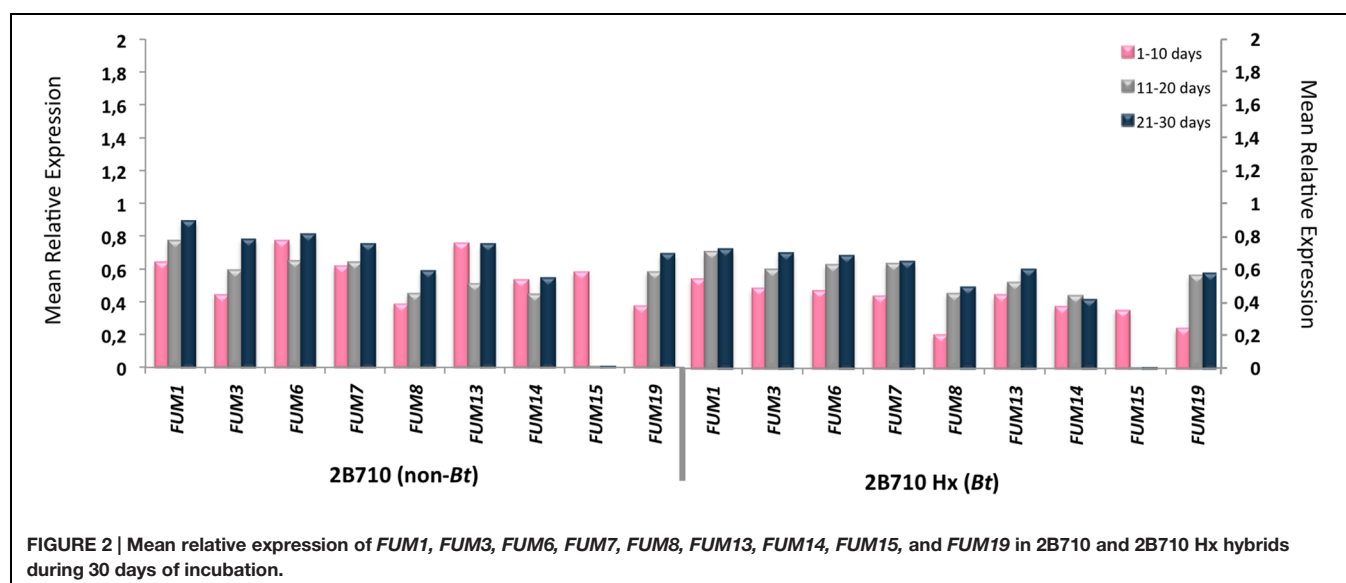
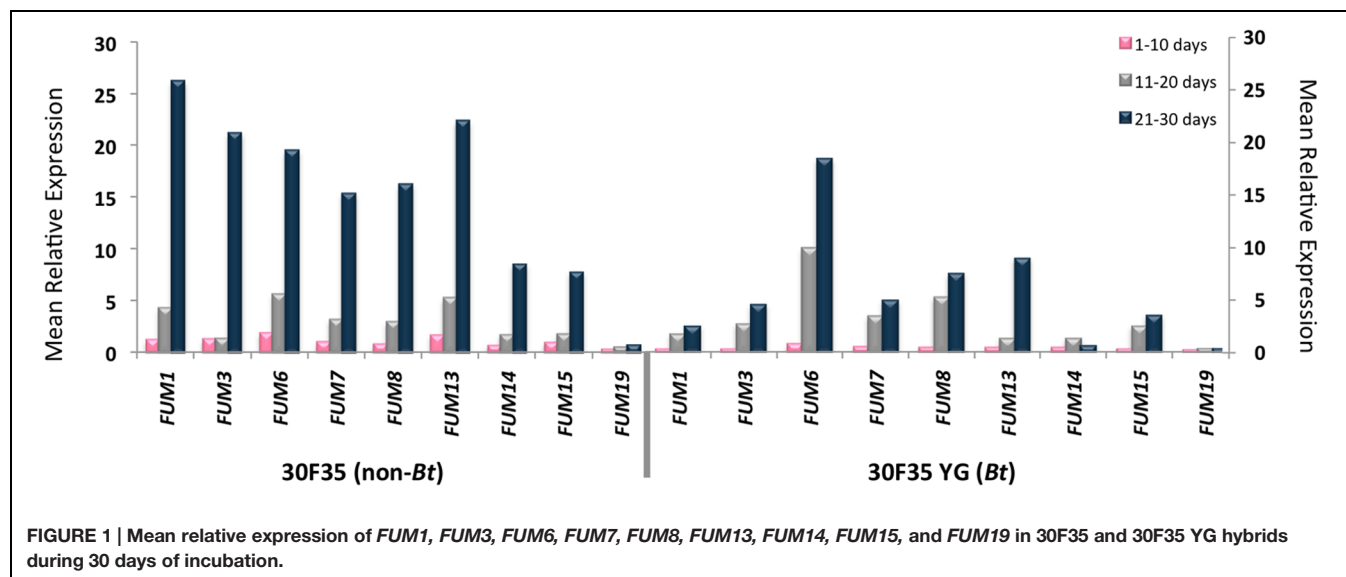
<sup>a</sup>Minimum fumonisin level for each 10, 20, and 30 days; <sup>b</sup>Maximum fumonisin level for each 10, 20, and 30 days; \*Mean ± standard error; <sup>c,d</sup>Differences of FB<sub>1</sub> production in 30F35 YG and 30F35, respectively ( $P < 0.05$ ); <sup>e,f</sup>Differences of FB<sub>1</sub> production in 2B710 Hx and 2B710, respectively ( $P < 0.05$ ).

The *Bt* hybrids 30F35 YG and 2B710 Hx were chosen due to their worldwide utilization for controlling insect pests. 30F35 YG expresses cry1Ab, an efficient protein for reducing injuries from lepidopteran pests' feeding (Bowers et al., 2014). Although cry1Ab is not highly effective against CEW (Bowers et al., 2013) and FAW, important lepidopteran insects found in Brazil (Farias et al., 2014), this protein has demonstrated efficacy in preventing insect injuries and therefore aiding in the reduction of fungal contamination and mycotoxin production (Munkvold, 2003; Ostry et al., 2010; Leslie and Logrieco, 2014).

2B710 Hx expresses cry1F protein, and it has been shown to be ineffective against CEW (Bowers et al., 2013), however, hybrids

expressing this protein have demonstrated temporary efficacy in controlling FAW in Brazil, since cry1F resistant populations of *S. frugiperda* have emerged from maize field where cry1F protein has been extensively used (Farias et al., 2014). This insect is native to the tropical regions of the western hemisphere from the United States to Argentina. The larvae feed primarily on maize leaf tissue, causing extensive defoliation (Farias et al., 2014); in addition, *S. frugiperda* can migrate to corn ear, causing damage to kernels (Gorman and Kang, 1991), creating infection sites for toxigenic fungi, such as *Fusarium* and *Aspergillus* (Munkvold, 2003, 2014).

Complementary research conducted by Michelotto et al. (2011), using the field and maize samples corresponding to those



used in this study, has demonstrated that the damage caused by *S. frugiperda* was significantly lower in 30F35 YG and 2B710 Hx when compared to their respective isogenic hybrids from sowing to harvest. This may explain the pattern of fumonisin production in the samples under controlled conditions, since damaged kernels can promote *F. verticillioides* invasion and therefore facilitate fumonisin production in non-*Bt* maize. In fact, insects are able to cause injuries to the external protection of grains and plant tissues in the field, allowing the fungal hyphae to readily penetrate, have access to nutrients and produce mycotoxins (Wu et al., 2004; Wu, 2006; Jouany, 2007). Since *Bt* maize reduces insect damage and the risk of fungal contamination, it could indirectly control mycotoxin accumulation during storage. However, it is important to emphasize that a deeper investigation regarding the differences between *Bt* and non-*Bt* maize in *F. verticillioides* contamination and fumonisin production as well

as the use of different cry proteins to effectively control insect pests are warranted to aid in the management strategies and to better understand the correlation between insect pest control and fungal invasion/mycotoxin production in maize.

The current study also compared the fumonisin production by *F. verticillioides* with FUM1, FUM3, FUM6, FUM7, FUM8, FUM13, FUM14, FUM15, and FUM19 gene expression for each of the studied hybrids, and also compared the FUM gene expression between *Bt* and non-*Bt* hybrids.

FUM genes were expressed by *F. verticillioides* in all of the corn samples, with an indication of lower gene expression for some of the studied genes in the *Bt* hybrids. There was a weak correlation between FB<sub>1</sub> and FB<sub>2</sub> production and the relative expression of some of the studied FUM genes were observed in 30F35 YG. Among these genes, FUM1, FUM14, and FUM19 were better correlated with fumonisin production. Interestingly, it

was observed differences in the expression of these same genes in *Bt* and non-*Bt* hybrids.

Studies have shown a correlation between some of the *FUM* genes and fumonisin production (Lopez-Errasquín et al., 2007; Jurado et al., 2010; Rocha et al., 2011; Lazzaro et al., 2012; Medina et al., 2013). *FUM19* encodes a protein that is highly similar to the ATP-binding cassette of multidrug resistance transporters, which act as efflux pumps to reduce cellular concentrations of toxins and thereby confer protection (Desjardins, 2006). Although Lopez-Errasquín et al. (2007), Jurado et al. (2010) and Rocha et al. (2011), have found a positive correlation between *FUM19* and fumonisin production, our results have shown a weak association between this gene expression and fumonisin production in 30F35 YG hybrid. *FUM14* also had a weak association with fumonisin production; this gene encodes a protein involved in the formation of tricarballic ester function. Indeed, a previous study has demonstrated that *FUM14*, as well as *FUM1* and *FUM19* exhibit a significant effect on the synthesis of FB<sub>1</sub> and FB<sub>2</sub> (Medina et al., 2013). Further studies regarding the expression of these genes should be conducted to verify whether or not they could be useful for diagnostics.

It is relevant to emphasize that previous studies have used synthetic media to produce fumonisins and analyze *FUM* gene expression (Lopez-Errasquín et al., 2007; Jurado et al., 2010; Rocha et al., 2011; Lazzaro et al., 2012; Medina et al., 2013), which increases the quality and concentration of RNA. Fungal RNA extraction from maize can be limited, once kernel consists of 90% starch and 10% proteins (Gibbon and Larkins, 2005). Even though the experimental Petri dishes containing maize grains were covered in *F. verticillioides* mycelium, the final RNA consisted of plant and fungal RNA, which likely decreased the performance of qPCR reactions, thereby interfering in the correlation between *FUM* gene cluster and fumonisin production. The rationale for isolating fungal RNA from maize grains is due to the fact that *F. verticillioides* can persist in undamaged plants and produce fumonisins; thus, identifying an adequate genetic marker for assessing fumonisin production during interaction between *F. verticillioides* and symptomless plants could aid in the prevention of fumonisin accumulation in the field and storage (Presello et al., 2007; Duncan and Howard, 2010; Bowers et al., 2013).

## CONCLUSION

Non-*Bt* maize samples 30F35 and 2B710 presented damaged kernels caused by insects in the field. In this study, wounded grains have possibly facilitated *F. verticillioides* penetration, leading to higher fumonisin production in non-*Bt* hybrids during the period of incubation due to the increase of fungal biomass. Insects cause physical damage, disruption of nutrients and grain injuries to several crops. Management

of insects through the use of *Bt* maize can greatly reduce insect damage, thus indirectly controlling fungal and mycotoxin contamination in the field and the accumulation of mycotoxins during storage. Despite the known effect of *Bt* maize on fumonisin content in the field, due to the control of insect pests, there is a lack of information regarding the accumulation of mycotoxins in *Bt* maize during a post-harvest period. Highlighting that, further investigations using different *Bt* hybrids could possibly aid in the controlling strategies. The gene expression analyses demonstrated that further studies should be conducted to determine an acceptable qualitative genetic marker for identifying fumonisin production in early stages of infection by *F. verticillioides* in maize.

## AUTHOR CONTRIBUTIONS

LR, BC, and AD conceived the study. AD and MM provided the maize samples. MM performed field sampling and initial tests for grain damage. VB and LA conducted fumonisin analysis. FF-C conducted gene expression tests. GP performed statistical analyses. LR and GP analyzed the overall data for the manuscript. LR wrote the manuscript. All authors approved the final version of the manuscript.

## FUNDING

This research was funded by FAPESP (Fundação de Amparo a Pesquisa do Estado de São Paulo).

## ACKNOWLEDGMENTS

The authors would like to thank FAPESP (Fundação de Amparo a Pesquisa do Estado de São Paulo) for providing financial support. We also thank the group of Instituto Agrônomo de Campinas, Cruzália - SP for technical assistance and suggestions during field sampling. We are grateful to Tatiana Alves dos Reis and Sabina Moser Tralamazza for technical assistance with HPLC method validation and gene expression analysis, respectively.

## SUPPLEMENTARY MATERIAL

The Supplementary Material for this article can be found online at: <http://journal.frontiersin.org/article/10.3389/fmicb.2015.01503>

**FIGURE S1 | Optimization of qPCR reactions. (A) *FUM1*, (B) *FUM3*, (C) *FUM6*, (D) *FUM7*, (E) *FUM8*, (F) *FUM13*, (G) *FUM14*, (H) *FUM15*, and (I) *FUM19* and reference genes [(J) *TUB* and (K) *CALM*].** Melt- curve, slope,  $r^2$ , and efficiency results for each of the studied genes. Curves were generated by four dilutions of total fungal RNA (10-fold dilutions for each point of the curve) extracted from maize grains contaminated with *Fusarium verticillioides* mycelium. Five replicates were tested for each dilution, and the slope,  $r^2$  and efficiencies were calculated.



## REFERENCES

- Abbas, H. K., Shier, W. T., and Cartwright, R. D. (2011). Effect of temperature, rainfall and planting date on aflatoxin and fumonisin contamination in commercial Bt and non-Bt corn hybrids. *Phytoprotection* 88, 41–50. doi: 10.7202/018054ar
- Berjak, P. (1984). Report of seed storage committee working group on the effects of storage fungi on seed viability. *Seed Sci. Technol.* 12, 233–253.
- Bolja, R. S., Cerny, R. L., Proctor, R. H., and Liangcheng, D. (2004). Determining the biosynthetic sequence in the early steps of the fumonisin pathway by use of three gene-disruption mutants of *Fusarium verticillioides*. *J. Agric. Food Chem.* 52, 2855–2860. doi: 10.1021/jf035429z
- Bowers, E. L., Hellmich, R. L., and Munkvold, G. P. (2013). Vip3Aa and Cry1Ab proteins in maize reduce *Fusarium* ear rot and fumonisins by deterring kernel injury from multiple Lepidopteran pests. *World Mycotoxin J.* 6, 127–135. doi: 10.3920/WMJ2012.1510
- Bowers, E. L., Hellmich, R. L., and Munkvold, G. P. (2014). Comparison of fumonisin contamination using HPLC and ELISA methods in Bt and near-isogenic maize hybrids infested with European corn borer or western bean cutworm. *J. Agric. Food Chem.* 62, 6463–6472. doi: 10.1021/jf5011897
- Brown, D. W., Butchko, R. A. E., Busman, M., and Proctor, R. H. (2007). The *Fusarium verticillioides* FUM gene cluster encodes a Zn(II)2Cys6 protein that affects FUM gene expression and fumonisin production. *Eukaryot. Cell* 6, 1210–1218. doi: 10.1128/EC.00400-06
- Bustin, S. A., Benes, V., Garson, J. A., Hellems, J., Huggett, J., Kubista, M., et al. (2009). The MIQE guidelines: minimum information for publication of quantitative real-time PCR experiments. *Clin. Chem.* 55, 611–622. doi: 10.1373/clinchem.2008.112797
- de la Campa, R., Hooker, D. C., Miller, J. D., Schaafsma, A. W., and Hammond, B. G. (2005). Modeling effects of environment, insect damage, and Bt genotypes on fumonisin accumulation in maize in Argentina and the Philippines. *Mycopathologia* 159, 539–552.
- Delp, R. B., Stewell, L. J., and Marois, J. J. (1986). Evaluation of field sampling techniques for estimation of disease incidence. *Phytopathology* 76, 1299–1305. doi: 10.1094/Phyto-76-1299
- Desjardins, A. E. (2006). *Fusarium Mycotoxins. Chemistry, Genetics and Biology*. St Paul, MN: APS Press.
- Dowd, P. F. (2000). Indirect reduction of ear molds and associated mycotoxins in *Bacillus thuringiensis* corn under controlled and open field conditions: utility and limitations. *J. Econ. Entomol.* 93, 1669–1679. doi: 10.1603/0022-0493-93.6.1669
- Duncan, K. E., and Howard, R. J. (2010). Biology of maize kernel infection by *Fusarium verticillioides*. *Mol. Plant Microbe Interact.* 23, 6–16. doi: 10.1094/MPMI-23-1-0006
- Dunn, P. K., and Smyth, G. K. (1996). Randomized quantile residuals. *J. Comput. Graph. Stat.* 5, 236–244. doi: 10.1080/10618600.1996.10474708
- Duvick, J. (2001). Prospects for reducing fumonisin contamination of maize through genetic modification. *Environ. Health Perspect.* 109, 337–342. doi: 10.1289/ehp.01109s2337
- European Commission Regulation [EC] (2007). No. 1126/2007 of 28 September 2007 amending regulation (EC) No. 1881/2006. setting maximum levels for certain contaminants in foodstuffs as regards fusarium toxins in maize and maize products. *J. Eur. Union* 255, 14–17.
- European Commission Regulation [EC] (2013). No. 12571/2013 of 19 of November 2013. *Guidance Document on Analytical Quality Control and Validation Procedures for Pesticide Residues Analysis in Food and Feed*. Brussels: European Commission, Health and Consumer Protection Directorate-General.
- European Network of Gmo Laboratories [ENGL] (2008). *Definition of Minimum Performance Requirements for Analytical Methods of GMO Testing*. Available at: <http://gmo-crl.jrc.ec.europa.eu>
- Farias, J. R., Andow, D. A., Horikoshi, R. J., Sorgatto, R. J., Fresia, P., dos Santos, A. C., et al. (2014). Field-evolved resistance to Cry1F maize by *Sporodoptera frugiperda* (Lepidoptera: Noctuidae) in Brazil. *Crop Prot.* 64, 150–158. doi: 10.1603/EC14190
- Ferreira, F. L., Aquino, S., Ribeiro, D. H. B., and Correa, B. (2007). Effects of gamma radiation on maize samples contaminated with *Fusarium verticillioides*. *Appl. Radiat. Isot.* 65, 927–933. doi: 10.1016/j.apradiso.2007.03.011
- Ferreira-Castro, F. L., Potenza, M. R., Rocha, L. O., and Correa, B. (2012). Interaction between toxigenic fungi and weevils in corn grain samples. *Food Control* 26, 594–600. doi: 10.1016/j.foodcont.2012.02.016
- Food and Drug Administration [FDA] (2001). *Background Paper in Support of Fumonisin Levels in Corn and Corn Products Intended for Human Consumption*. College Park, MD: US Food and Drug Administration, Center for Food Safety and Applied Nutrition.
- Foresti, J., Bernardi, O., Zart, M., and Garcia, M. S. (2013). Oviposition behavior of *Helicoverpa zea* (Boddie, 1850) (Lepidoptera: Noctuidae) in corn seed and simulation of control. *Rev. Bras. Milho Sorgo* 12, 78–84. doi: 10.18512/1980-6477/rbms.v12n1p78-84
- Gibbon, B. C., and Larkins, B. A. (2005). Molecular genetic approaches to developing quality protein maize. *Trends Genet.* 21, 227–233. doi: 10.1016/j.tig.2005.02.009
- Gibbons, J. D., and Chakraborti, S. (2003). *Nonparametric Statistical Inference*. New York, NY: Marcel Dekker.
- Gizinger, D. G. (2002). Gene quantification using real-time quantitative PCR: an emerging technology hits the mainstream. *Exp. Hematol.* 30, 503–512. doi: 10.1016/S0301-472X(02)00806-8
- Gorman, D. P., and Kang, M. S. (1991). Preharvest aflatoxin contamination in maize: resistance and genetics. *Plant Breed.* 107, 1–10. doi: 10.1111/j.1439-0523.1991.tb00522.x
- Gradziel, T. M., and Wang, D. (1994). Susceptibility of California almond cultivars to aflatoxigenic *Aspergillus flavus*. *HortScience* 29, 33–35.
- Imathiu, S. M., Ray, R. V., Back, M., Hare, M. C., and Edwards, S. G. (2008). *Fusarium langsethiae* pathogenicity and aggressiveness towards oats and wheat in wounded and unwounded in vitro detached leaf assays. *Eur. J. Plant Pathol.* 124, 117–126. doi: 10.1007/s10658-008-9398-7
- Jouany, J. P. (2007). Methods for preventing decontamination and minimizing the toxicity of mycotoxins in feeds. *Anim. Feed Sci. Technol.* 137, 342–362. doi: 10.1016/j.anifeedsci.2007.06.009
- Jurado, M., Marín, P., Calleja, C., Moretti, A., Vázquez, C., and González-Jaén, M. T. (2010). Genetic variability and fumonisin production by *Fusarium proliferatum*. *Food Microbiol.* 27, 50–57. doi: 10.1016/j.fm.2009.08.001
- Lazzaro, I., Susca, A., Mulè, G., Ritieni, A., Ferracane, R., Marocco, A., et al. (2012). Effects of temperature and water activity on FUM2 and FUM21 gene expression and fumonisin B1 production in *Fusarium verticillioides*. *Eur. J. Plant Pathol.* 134, 685–695. doi: 10.1007/s10658-012-0045-y
- Leslie, K. F., and Logrieco, A. (2014). *Mycotoxin Reduction in Grain Chains*. Iowa City, IA: John Wiley and Sons, Inc.
- Lopez-Errasquin, E., Vazquez, C., Jimenez, M., and Gonzalez-Jaen, M. T. (2007). Real-Time RT-PCR assay to quantify the expression of FUM1 and FUM19 genes from the fumonisin-producing *Fusarium verticillioides*. *J. Microbiol. Methods* 68, 312–317. doi: 10.1016/j.mimet.2006.09.007
- Mahoney, M. E., and Rodrigues, S. B. (1996). Aflatoxin variability in pistachios. *Appl. Environ. Microbiol.* 62, 1197–1202.
- Marasas, W. F. O., Riley, R. T., Hendricks, K. A., Stevens, V. L., Sadler, T. W., Gelineau-Van Waes, J., et al. (2004). Fumonisin disrupt sphingolipid metabolism, folate transport, and neural tube development in embryo culture and in vivo: a potential risk factor for human neural tube defects among populations consuming fumonisin-contaminated maize. *J. Nutr.* 134, 711–716.
- Medina, A., and Magan, N. (2010). Comparisons of water activity and temperature impacts on growth of *Fusarium langsethiae* strains from northern Europe on oat-base media. *Int. J. Food Microbiol.* 142, 365–369. doi: 10.1016/j.ijfoodmicro.2010.07.021
- Medina, A., Schmidt-Heydt, M., Cárdenas-Chávez, L., Parra, R., Geisen, R., and Magan, N. (2013). Integrating toxin gene expression, growth and fumonisin B1 and B2 production by a strain of *Fusarium verticillioides* under different environmental factors. *J. R. Soc. Interface* 10, 1–12. doi: 10.1098/rsif.2013.0320
- Michelotto, M. D., Pereira, A. D., Finoto, E. L., and Freitas, R. S. (2011). Controle de pragas em híbridos de milho geneticamente modificados. *Grandes Cult.* 145, 36–38.
- Mohammadi, M. M., and Hokmabadi, H. (2010). Study on the effect of pistachio testa on the reduction of *Aspergillus flavus* growth and aflatoxin B1 production in kernels of different pistachio cultivars. *Aust. J. Crop Sci.* 4, 744–749.
- Munkvold, G. P. (2003). Cultural and genetic approaches to managing mycotoxins in maize. *Annu. Rev. Phytopathol.* 41, 99–116. doi: 10.1146/annurev.phyto.41.052002.095510

- Munkvold, G. P. (2014). "Crop management practices to minimize the risk of mycotoxins contamination in temperate-zone maize," in *Mycotoxin Reduction in Grain Chains*, eds J. F. Leslie and A. Logrieco (Iowa City, IA: Wiley Blackwell), 59–75.
- Munkvold, G. P., and Hellmich, R. L. (1999). Comparison of fumonisin concentrations in kernels of transgenic Bt maize hybrids and nontransgenic hybrids. *Plant Dis.* 83, 130–138. doi: 10.1007/BF03036703
- Munkvold, G. P., Hellmich, R. L., and Rice, L. G. (1999). Comparison of fumonisin concentrations in kernels of transgenic Bt maize hybrids and non-transgenic hybrids. *Plant Dis.* 83, 130–138. doi: 10.1094/PDIS.1999.83.2.130
- Munkvold, G. P., Hellmich, R. L., and Showers, W. B. (1997). Reduced *Fusarium* ear rot and symptomless infection in kernels of maize genetically engineered for European corn borer resistance. *Phytopathology* 87, 1071–1077. doi: 10.1094/PHYTO.1997.87.10.1071
- Oren, L., Ezrati, S., Cohen, D., and Sharon, A. (2003). Early events in the *Fusarium verticillioides*–maize interaction characterized by using a green fluorescent protein-expressing transgenic isolate. *Appl. Environ. Microbiol.* 69, 1695–1701. doi: 10.1128/AEM.69.3.1695-1701.2003
- Ostry, V., Ovesna, J., Skarkova, J., Pouchova, V., and Ruprich, J. (2010). A review on comparative data concerning *Fusarium* mycotoxins in Bt maize and non-Bt isogenic maize. *Mycotoxin Res.* 26, 141–145. doi: 10.1007/s12550-010-0056-5
- Parsons, M. W., and Munkvold, G. P. (2012). Effects of planting date and environmental factors on *Fusarium* ear rot symptoms and fumonisin B1 accumulation in maize grown in six North American locations. *Plant Pathol.* 61, 1130–1142. doi: 10.1111/j.1365-3059.2011.02590.x
- Pazzi, F., Lener, M., Colombo, L., and Monastera, G. (2006). Bt maize and mycotoxins: the current state of research. *Ann. Microbiol.* 56, 223–230. doi: 10.1007/BF03175009
- Pfaffl, M. W. (2001). A new mathematical model for relative quantification in real time RT-PCR. *Nucleic Acids Res.* 29, 2002–2007. doi: 10.1093/nar/29.9.e45
- Presello, D. A., Iglesias, J., Botta, G., and Eyherabide, G. H. (2007). Severity of *Fusarium* ear rot and concentration of fumonisin in grain of Argentinian maize hybrids. *Crop Prot.* 26, 852–855. doi: 10.1094/PHYTO.2004.94.3.251
- Proctor, R. H., Brown, D. W., Plattner, R. D., and Desjardins, A. E. (2003). Co-expression of 15 contiguous genes delineates a fumonisin biosynthetic gene cluster in *Gibberella moniliformis*. *Fungal Genet. Biol.* 38, 237–249. doi: 10.1016/S1087-1845(02)00525-X
- Proctor, R. H., Desjardins, A. E., Plattner, R. D., and Hohn, T. M. (1999). A polyketide synthase gene required for biosynthesis of fumonisin mycotoxins in *Gibberella fujikuroi* mating population A. *Fungal Genet. Biol.* 7, 100–112. doi: 10.1006/fgbi.1999.1141
- Proctor, R. H., Van Hove, F., Susca, A., Stea, G., Busman, M., Van der Lee, T., et al. (2013). Birth, death and horizontal gene transfer of the fumonisin biosynthetic gene cluster during the evolutionary diversification of *Fusarium*. *Mol. Microbiol.* 90, 290–306. doi: 10.1111/mmi.12362
- Raymaekers, M., Smets, R., Maes, B., and Cartuyvels, R. (2009). Checklist for optimization and validation of real-time PCR assays. *J. Clin. Lab. Anal.* 23, 145–151. doi: 10.1002/jcla.20307
- Rigby, R. A., and Stasinopoulos, D. M. (2005). Generalized additive models for location, scale and shape. *J. R. Stat. Soc. C Appl. Stat.* 54, 507–554. doi: 10.1111/j.1467-9876.2005.00510.x
- Rocha, L. O., Reis, G. M., da Silva, V. N., Braghini, R., Teixeira, M. M. G., and Corrêa, B. (2011). Molecular characterization and fumonisin production by *Fusarium verticillioides* isolated from corn grains of different geographic origins in Brazil. *Int. J. Food Microbiol.* 145, 9–21. doi: 10.1016/j.jfoodmicro.2010.11.001
- Ross, P. F., Nelson, P. E., Richard, J. L., Plattner, R. D., Rice, L. G., Osweiler, G. D., et al. (1990). Production of fumonisin by *Fusarium verticillioides* and *Fusarium proliferatum* isolates associated with equine leukoencephalomalacia and pulmonary oedema syndrome swine. *Appl. Environ. Microbiol.* 56, 3224–3226.
- Schatzmayer, G., and Streit, E. (2013). Global occurrence of mycotoxins in the food and feed chain: facts and figures. *World Mycotoxin J.* 6, 213–222. doi: 10.3920/WMJ2013.1572
- Schmittgen, T. D., and Livak, K. J. (2008). Analyzing real-time PCR data by the comparative Ct method. *Nat. Protoc.* 3, 1101–1108. doi: 10.1038/nprot.2008.73
- Sinha, A. K. (1994). "The impact of insect pests on aflatoxin contamination of stored wheat and maize," in *Proceedings of the 6th International Working Conference on Stored-Product Protection*, eds E. Highley, E. J. Wright, H. J. Banks, and B. R. Champ (Wallingford: CAB International), 1059–1063.
- Stepien, L., Koczyk, G., and Waskiewicz, A. (2011). Genetic and phenotypic variation of *Fusarium proliferatum* isolates from different host species. *J. Appl. Genet.* 52, 487–496. doi: 10.1007/s13353-011-0059-8
- Sydenham, E. W., Thiel, P. G., Marasas, W. F. O., Shephard, G. S., Schalkwyk, D. J., and Koch, K. R. (1990). Natural occurrence of some *Fusarium* mycotoxins in corn from low and high esophageal cancer prevalence areas of the Transkei, Southern Africa. *J. Agric. Food Chem.* 38, 1900–1903. doi: 10.1021/jf00100a004
- United States Department of Agriculture [USDA] (2014). *World Agricultural Supply and Demand Estimates*. Available at: [www.usda.gov/oce/commodity/wasde/latest.pdf](http://www.usda.gov/oce/commodity/wasde/latest.pdf)
- Visconti, A., Solfrizzo, M., and Girolamo, A. (2001). Determination of fumonisins B1 and B2 in corn and corn flakes by liquid chromatography with immunoaffinity column cleanup: collaborative study. *J. AOAC Int.* 84, 1828–1837.
- Visentin, I., Tamiatti, G., Valentino, D., Portis, E., Karlovsky, P., Moretti, A., et al. (2009). The ITS region as a taxonomic discriminator between *Fusarium verticillioides* and *Fusarium proliferatum*. *Mycol. Res.* 113, 1137–1145. doi: 10.1016/j.mycres.2009.07.011
- Wallin, J. R. (1986). Production of aflatoxin in wounded and whole maize kernels by *Aspergillus flavus*. *Plant Dis.* 70, 429–430. doi: 10.1094/PD-70-429
- Winston, P. W., and Bates, D. H. (1960). Saturated solutions for the control of humidity in biological research. *Ecology* 41, 232–236. doi: 10.2307/1931961
- Wu, F. (2006). Mycotoxin reduction in Bt corn: potential economic, health, and regulatory impacts. *Transgenic Res.* 15, 277–289. doi: 10.1007/s11248-005-5237-1
- Wu, F., Miller, J. D., and Casman, E. A. (2004). Bt corn and mycotoxin reduction: economic impacts in the United States and the developing world. *J. Toxicol. Toxin Rev.* 23, 397–424. doi: 10.1081/TXR-200027872
- Yan, K., and Dickman, M. B. (1996). Isolation of a 3-tubulin gene from *Fusarium moniliforme* that confers cold-sensitive benomyl resistance. *Appl. Environ. Microbiol.* 62, 3053–3056.

**Conflict of Interest Statement:** The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

Copyright © 2016 Rocha, Barroso, Andrade, Pereira, Ferreira-Castro, Duarte, Michelotto and Correa. This is an open-access article distributed under the terms of the Creative Commons Attribution License (CC BY). The use, distribution or reproduction in other forums is permitted, provided the original author(s) or licensor are credited and that the original publication in this journal is cited, in accordance with accepted academic practice. No use, distribution or reproduction is permitted which does not comply with these terms.